

Big Data Analytics

Session 11

Predicting Algae Bloom

Problem Description

- Harmful algae in rivers
 - A serious ecological problem
 - Strong impact on
 - river life forms and
 - water quality
- Objectives:
 - Monitor and perform an early forecast of algae blooms
 - to improve the quality of rivers
 - chemical monitoring is cheaper and easily automated than biological analysis (microscopic examination)
 - Provide a better understanding of the factors influencing the algae frequencies



Data Collection



- Several water samples were collected in different European rivers at different times during a period of approximately 1 year.
- For each water sample,
 - different chemical properties were measured, as well as
 - the frequency of several harmful algae
- Some related characteristics were stored
 - the season of the year
 - the river size
 - the river speed
- Data was collected in the context of the ERUDIT research Network
 - available in the UCI machine learning repository
 - <http://archive.ics.uci.edu/ml/datasets/Coil+1999+Competition+Data>

Data Description



- Two main datasets:
 - Training dataset
 - 200 observations
 - 11 predictors
 - Nominal (3): season, size, speed
 - Numerical (8): different chemical parameters measured in the water samples
 - » Maximum pH value, Minimum value of O₂ (Oxygen)
 - » Mean value of Cl, NO₃⁻, NH₄⁺, PO₄³⁻, PO₄, chlorophyll
 - 7 responses
 - Seven frequency numbers of different harmful algae found in respective sample
 - Test dataset
 - 140 observations
 - 11 predictors
 - no responses

Goal: to predict the frequency of the seven algae for these 140 water samples

Load the Data into R



- Download the data (in .txt form) to your working directory (getwd()) from <http://www.dcc.fc.up.pt/~ltorgo/DataMiningWithR/datasets2.html> or <https://archive.ics.uci.edu/ml/machine-learning-databases/coil-mld/coil.html>
 - Analysis.txt**: training data; **Eval.txt**: test data

```
algae <- read.table('Analysis.txt', header=F, dec='.',  
                     col.names=c('season','size','speed','mxPH','mnO2','Cl','NO3','NH4','oPO4',  
                     'PO4','Chla','a1','a2','a3','a4','a5','a6','a7'),  
                     na.strings=c('XXXXXXX'))
```

#header=F: indicates that the file to be read does not include a first line with variable names

#dec='.': the numbers use '.' to separate decimal places (e.g., 34.2)

#na.strings: unknown values are represented by XXXXXX

or library(DMwR)

or algae <- read.table('/Users/than/.../Analysis.txt', header=F, dec='.', ...)

/Users/than/.../ is the directory where the training data is stored.

```
> head(algae)  
  season size speed mxPH mnO2 Cl NO3 NH4 oPO4 PO4 Chla a1 a2 a3 a4 a5 a6 a7  
1 winter small medium 8.00 9.8 60.800 6.238 578.000 105.000 170.000 50.0 0.0 0.0 0.0 0.0 34.2 8.3 0.0  
2 spring small medium 8.35 8.0 57.750 1.288 370.000 428.750 558.750 1.3 1.4 7.6 4.8 1.9 6.7 0.0 2.1  
3 autumn small medium 8.10 11.4 40.020 5.330 346.667 125.667 187.057 15.6 3.3 53.6 1.9 0.0 0.0 0.0 9.7  
4 spring small medium 8.07 4.8 77.364 2.302 98.182 61.182 138.700 1.4 3.1 41.0 18.9 0.0 1.4 0.0 1.4  
5 autumn small medium 8.06 9.0 55.350 10.416 233.700 58.222 97.580 10.5 9.2 2.9 7.5 0.0 7.5 4.1 1.0  
6 winter small high 8.25 13.1 65.750 9.248 430.000 18.250 56.667 28.4 15.1 14.6 1.4 0.0 22.5 12.6 3.9
```

Descriptive Data Analysis

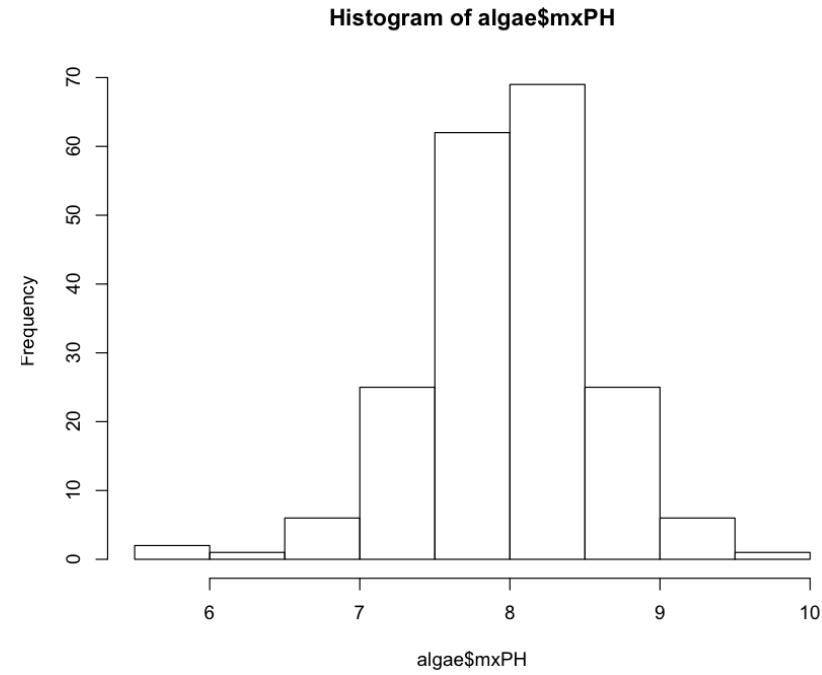
Data Visualisation and Summarisation

Data Visualisation and Summarisation

- Use summary(algae)
 - Notice the difference that nominal and numerical variables are presented
 - Nominal: frequency counts
 - Numerical: 5 number summary

```
season      size      speed      mxPH
autumn:40   large :45   high  :84   Min.   :5.600
spring:53   medium:84  low   :33   1st Qu.:7.700
summer:45   small :71   medium:83  Median  :8.060
winter:62

mno2          c1          NO3
Min.   : 1.500  Min.   : 0.222  Min.   : 0.050
1st Qu.: 7.725 1st Qu.: 10.981 1st Qu.: 1.296
Median  : 9.800 Median  : 32.730 Median  : 2.675
Mean    : 9.118 Mean   : 43.636 Mean   : 3.282
3rd Qu.:10.800 3rd Qu.: 57.824 3rd Qu.: 4.446
Max.   :13.400  Max.   :391.500 Max.   :45.650
NA's    :2        NA's   :10     NA's   :2
```



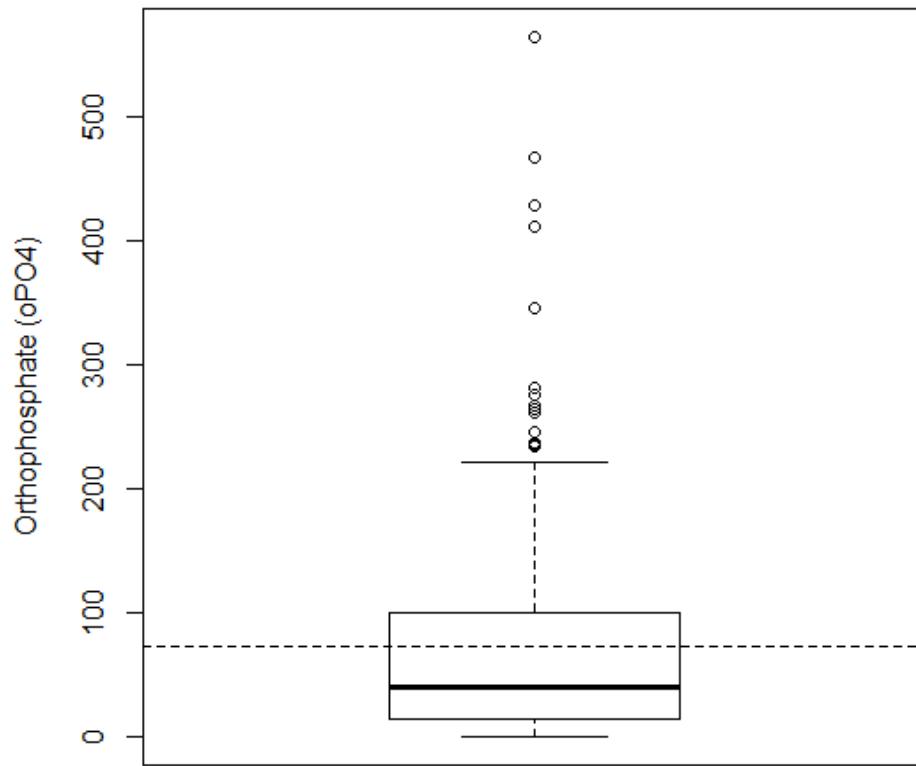
- Use graphs to check the shape of distribution

```
> hist(algae$mxPH) # the shape suggests that mxPH is nearly normal distributed
```

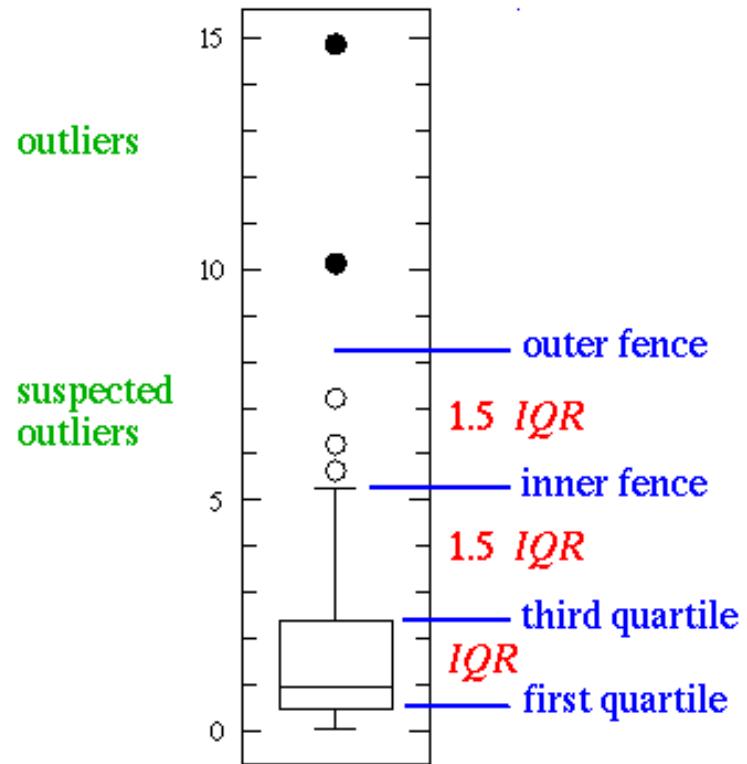
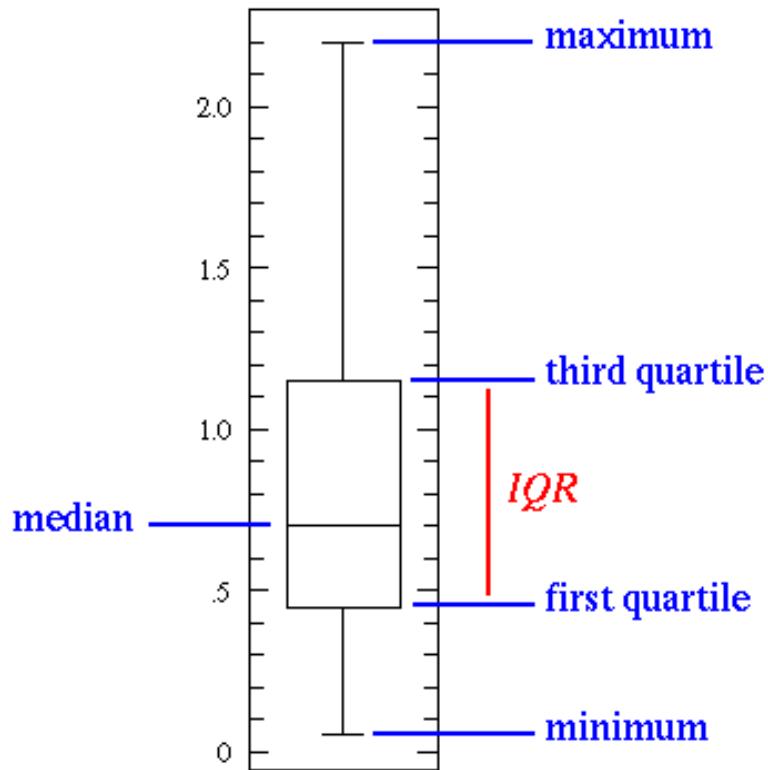
Data Visualisation and Summarisation

- Or boxplot

```
> boxplot(algae$oPO4, ylab='Orthophosphate (oPO4)')  
> abline(h=mean(algae$oPO4, na.rm=T), lty=2)
```

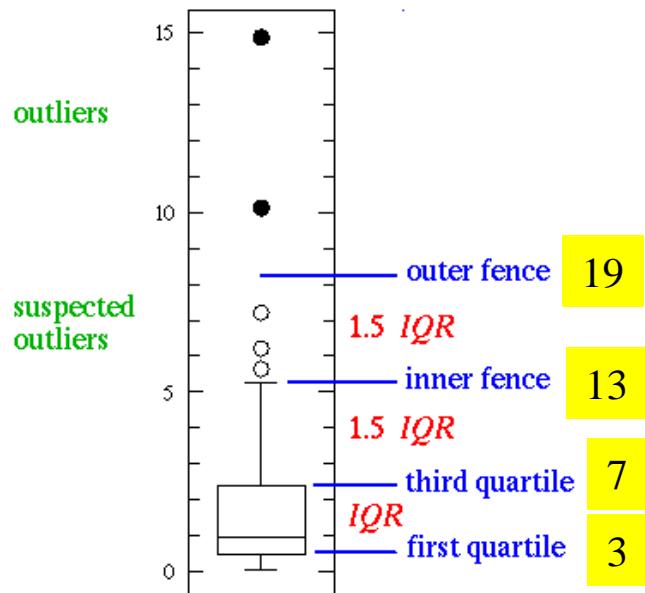


Boxplots and Outliers



In R, all the suspected outliers and outliers are unfilled circles.

Boxplots and Outliers



data is 1,2,3,4,5,6,7,8,x

Q1=3, Q2=5, Q3=7

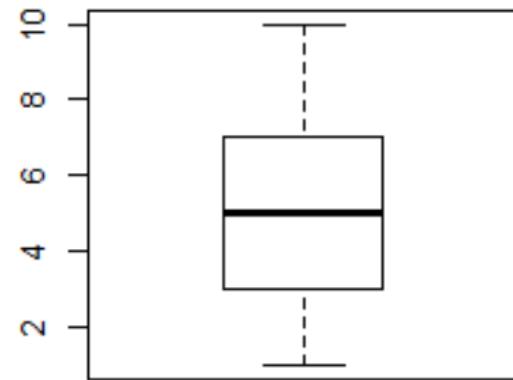
IQR = 7-3 = 4

1.5*IQR = 6

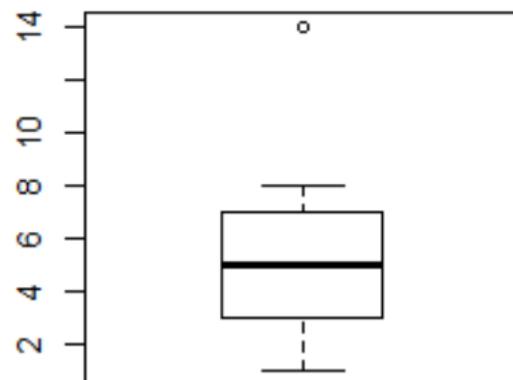
3*IQR = 12

inner fence = Q3+1.5*IQR = 13

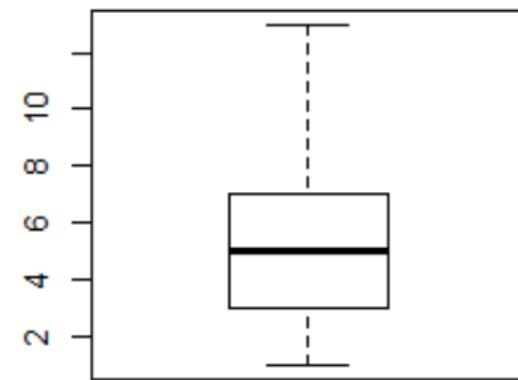
outer fence = Q3+3*IQR = 19



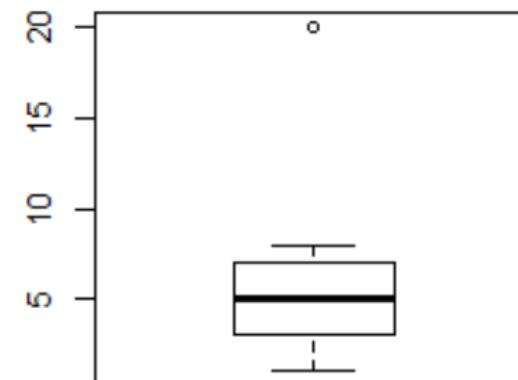
values1 = c(1:8, 10)



values3 = c(1:8, 14)



values2 = c(1:8, 13)



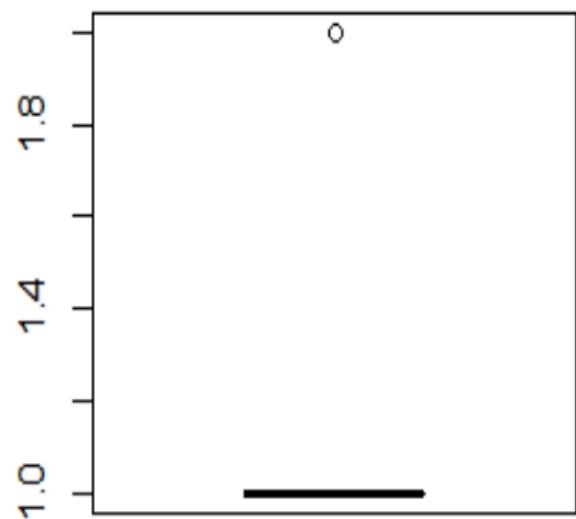
values4 = c(1:8, 20)

Outliers – A Remark



- Outliers are not necessarily "bad" data-points
- They may well be the most important, most information rich, part of the dataset
- Under no circumstances should they be automatically removed from the dataset
- Outliers may deserve special consideration
 - they may be the key to the phenomenon under study or the result of human blunders

rug and jitter



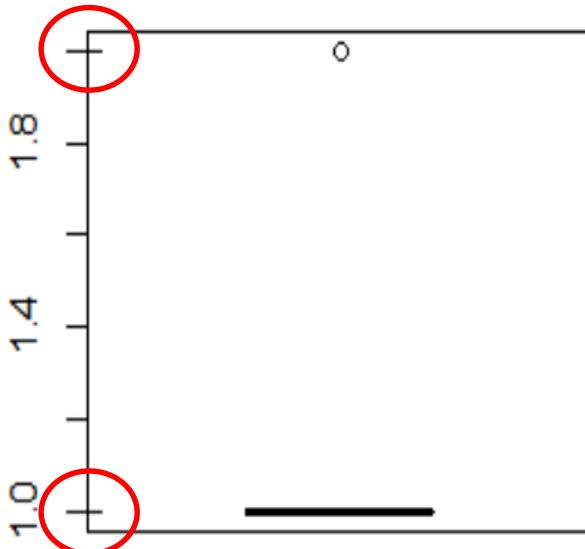
```
> values <- c(1,1,1,1,1,1,2)
```

```
> boxplot(values)
```

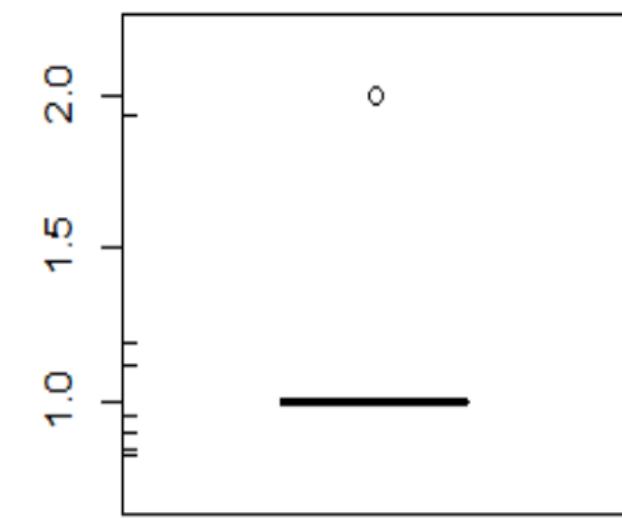
```
> set.seed(7)
```

```
> jitter(values)
```

```
[1] 1.1955637 0.9590982 0.8462791 0.8278995 0.8974998 1.1168042 1.9360249
```



```
> rug(values, side=2)
```



```
> values = c(1,1,1,1,1,1,2)
```

```
> boxplot(values, ylim=c(0.7,2.2))
```

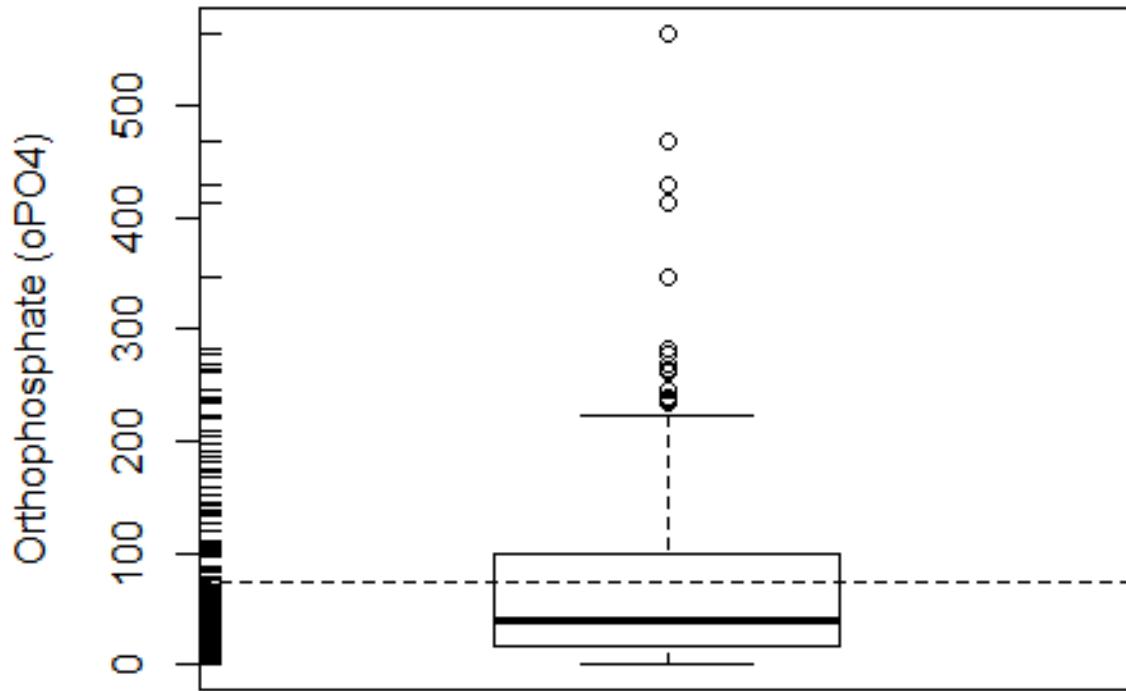
```
> set.seed(7)
```

```
> rug(jitter(values), side=2)
```

Data Visualisation and Summarisation

- Or boxplot

```
> boxplot(algae$oPO4, ylab='Orthophosphate (oPO4)')  
> abline(h=mean(algae$oPO4, na.rm=T), lty=2)  
> rug(jitter(algae$oPO4), side=2) #side=2 - left, 3-up, 4-right, 1-bottom
```



jitter:

Add a small amount of noise to a numeric vector.

rug:

Adds a set of tick marks along the base of a plot.

Data Visualisation and Summarisation



- Detect outliers with graphics

```
>plot(algae$NH4,xlab='')
```

```
>abline(h=mean(algae$NH4,na.rm=T),lty=1,col="red")
```

```
>abline(h=mean(algae$NH4,na.rm=T)+sd(algae$NH4,na.rm=T),lty=2,col="blue")
```

```
>abline(h=median(algae$NH4,na.rm=T),lty=3,col="green")
```

```
>identify(algae$NH4)
```

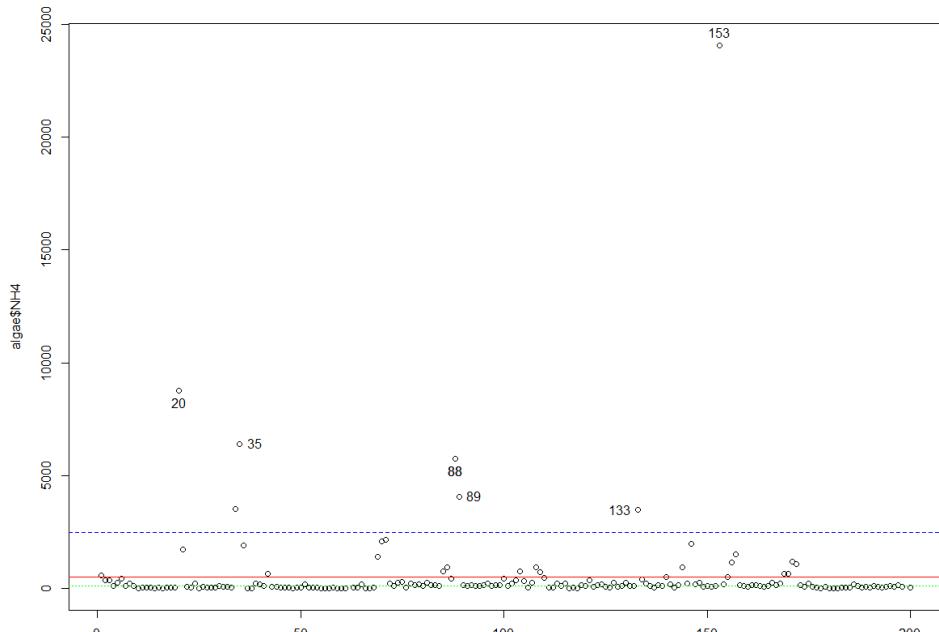
`identify` is interactive: when a user click on the plotted dots with the left mouse, the row number of that observation will be shown. Click right mouse to finish interaction.

`identify` might not work in RStudio, though.

Try the original R tool instead.

- Detect outliers without graphics

```
> algae[algae$NH4 >19000, ]
      season size speed mxPH mnO2 Cl       NO3
NA    <NA>   <NA> <NA>   NA   NA   NA   NA
153  autumn medium high  7.3 11.8 44.205 45.65
NA.1 <NA>   <NA> <NA>   NA   NA   NA   NA
```



Data Preprocessing

Dealing with Missing Values

Dealing With Unknown Values



- Unknown (missing) values
 - are common in real-world problems
 - may preclude the use of certain statistical learning approaches
 - E.g., randomForest(), mean()
- Solutions
 - Remove the cases with unknowns
 - Fill in the unknown values by exploring the most frequent value
 - Fill in the unknown values by exploring the correlations between variables
 - Fill in the unknown values by exploring the similarity between cases
 - Use tools that are able to handle these values

Removing the Obs. with Unknown Values



- Before removing them, check/count them first

```
> library(DMwR)
> data(algae)      # load fresh data again before we try different ways of dealing with unknown values
> algae[!complete.cases(algae),]    # check whether each obs is complete or not
   season   size   speed mxPH mnO2     Cl    NO3 NH4    oPO4    PO4 Chla   a1    a2    a3    a4    a5    a6    a7
28 autumn small  high  6.80 11.1 9.000 0.630  20  4.000    NA  2.70 30.3  1.9 0.0  0.0 2.1 1.4 2.1
38 spring  small  high  8.00    NA 1.450 0.810  10  2.500  3.000  0.30 75.8  0.0 0.0  0.0 0.0 0.0 0.0
48 winter small  low   NA    12.6 9.000 0.230  10  5.000  6.000  1.10 35.5  0.0 0.0  0.0 0.0 0.0 0.0
55 winter small  high  6.60 10.8    NA 3.245 10  1.000  6.500    NA 24.3 0.0 0.0  0.0 0.0 0.0 0.0
56 spring  small  medium 5.60 11.8    NA 2.220  5  1.000  1.000    NA 82.7 0.0 0.0  0.0 0.0 0.0 0.0
57 autumn small  medium 5.70 10.8    NA 2.550 10  1.000  4.000    NA 16.8 4.6 3.9 11.5 0.0 0.0 0.0
58 spring  small  high  6.60  9.5    NA 1.320 20  1.000  6.000    NA 46.8 0.0 0.0  28.8 0.0 0.0 0.0
59 summer  small  high  6.60 10.8    NA 2.640 10  2.000 11.000    NA 46.9 0.0 0.0 13.4 0.0 0.0 0.0
60 autumn small  medium 6.60 11.3    NA 4.170 10  1.000  6.000    NA 47.1 0.0 0.0  0.0 0.0 1.2 0.0
61 spring  small  medium 6.50 10.4    NA 5.970 10  2.000 14.000    NA 66.9 0.0 0.0  0.0 0.0 0.0 0.0
62 summer  small  medium 6.40    NA    NA    NA    NA 14.000    NA 19.4 0.0 0.0  2.0 0.0 3.9 1.7
63 autumn small  high  7.83 11.7 4.083 1.328 18  3.333  6.667    NA 14.4 0.0 0.0  0.0 0.0 0.0 0.0
116 winter medium high  9.70 10.8 0.222 0.406 10 22.444 10.111    NA 41.0 1.5 0.0  0.0 0.0 0.0 0.0
161 spring  large  low   9.00  5.8    NA 0.900 142 102.000 186.000 68.05 1.7 20.6 1.5  2.2 0.0 0.0 0.0
184 winter  large  high  8.00 10.9 9.055 0.825 40 21.083  56.091    NA 16.8 19.6 4.0  0.0 0.0 0.0 0.0
199 winter  large  medium 8.00  7.6    NA    NA    NA    NA    NA    NA 0.0 12.5 3.7  1.0 0.0 0.0 4.9
>
> nrow(algae[!complete.cases(algae),])
[1] 16
>
> algae <- na.omit(algae)
[1] 184
```

Removing the Obs. with Unknown Values



- Probably think twice before removing so many observations

```
> library(DMwR)
> data(algae) # load fresh data again before we try different ways of dealing with unknown values
> algae[!complete.cases(algae),] # check whether each observation is complete or not
   season size speed mxPH mnO2   Cl   NO3 NH4    OPO4    PO4 Chla   a1   a2   a3   a4   a5   a6   a7
28 autumn small  high 6.80 11.1 9.000 0.630  20  4.000     NA 2.70 30.3 1.9 0.0 0.0 2.1 1.4 2.1
38 spring  small  high 8.00   NA 1.450 0.810  10  2.500  3.000 0.30 75.8 0.0 0.0 0.0 0.0 0.0 0.0 0.0
48 winter  small   low  NA 12.6 9.000 0.230  10  5.000  6.000 1.10 35.5 0.0 0.0 0.0 0.0 0.0 0.0 0.0
55 winter  small  high 6.60 10.8   NA 3.245  10  1.000  6.500  NA 24.3 0.0 0.0 0.0 0.0 0.0 0.0 0.0
56 spring  small medium 5.60 11.8   NA 2.220   5  1.000  1.000  NA 82.7 0.0 0.0 0.0 0.0 0.0 0.0 0.0
57 autumn  small medium 5.70 10.8   NA 2.550  10  1.000  4.000  NA 16.8 4.6 3.9 11.5 0.0 0.0 0.0 0.0 0.0
58 spring  small  high 6.60  9.5   NA 1.320  20  1.000  6.000  NA 46.8 0.0 0.0 28.8 0.0 0.0 0.0 0.0 0.0
59 summer  small  high 6.60 10.8   NA 2.640  10  2.000 11.000  NA 46.9 0.0 0.0 13.4 0.0 0.0 0.0 0.0 0.0
60 autumn  small medium 6.60 11.3   NA 4.170  10  1.000  6.000  NA 47.1 0.0 0.0 0.0 0.0 0.0 1.2 0.0
61 spring  small medium 6.50 10.4   NA 5.970  10  2.000 14.000  NA 66.9 0.0 0.0 0.0 0.0 0.0 0.0 0.0 0.0
62 summer  small medium 6.40   NA    NA    NA     NA 14.000  NA 19.4 0.0 0.0 2.0 0.0 3.9 1.7
63 autumn  small  high 7.83 11.7 4.083 1.328  18  3.333  6.667  NA 14.4 0.0 0.0 0.0 0.0 0.0 0.0 0.0
116 winter  medium high 9.70 10.8 0.222 0.406  10 22.444 10.111  NA 41.0 1.5 0.0 0.0 0.0 0.0 0.0 0.0
161 spring  large   low 9.00  5.8   NA 0.900 142 102.000 186.000 68.05 1.7 20.6 1.5 2.2 0.0 0.0 0.0 0.0
184 winter  large  high 8.00 10.9 9.055 0.825  40 21.083 56.091  NA 16.8 19.6 4.0 0.0 0.0 0.0 0.0 0.0
199 winter  large medium 8.00  7.6    NA    NA     NA     NA  NA 0.0 12.5 3.7 1.0 0.0 0.0 0.0 4.9
>
> manyNAs(algae) # returns the row numbers that have more than 20% of the columns with an NA. In this case, 18*20% = 3.6 columns.
[1] 62 199
> algae <- algae[-c(62, 199), ]
> algae <- algae[-manyNAs(algae), ] # the last two commands have the same effect
```

Filling with Most Frequent Values



- Several alternatives can be chosen, with different trade-offs between
 - the level of approximation, and
 - the computational complexity of the method
- First alternative (simplest and fastest)
 - Use some statistics of centrality to fill in the unknown values
 - mean, median, mode, etc
 - choose mean if the distribution is nearly normal
 - choose median if not
 - For example,

```
season    size    speed  mxPH mnO2      Cl      NO3 NH4      oPO4      PO4     Chla     a1      a2      a3      a4      a5      a6      a7
48  winter   small    low    NA 12.6 9.000 0.230  10  5.000  6.000  1.10 35.5  0.0  0.0  0.0  0.0  0.0  0.0  0.0
```

This method is simple, fast, thus appealing for large dataset. However, it may introduce a large bias in the data.

Recall that the mxPH is nearly normal distributed, we could use its mean value to fill in the hole.

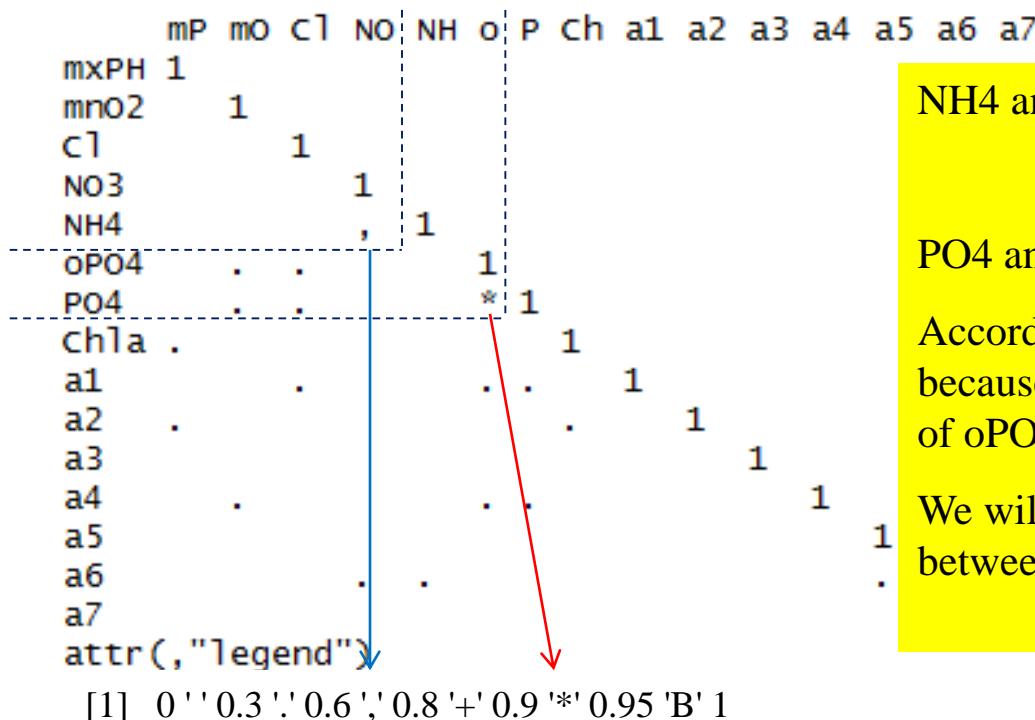
```
>algae[48, 'mxPH'] <- mean(algae$mxPH, na.rm=T)
#calculate the mean of the mxPH column while ignoring any NA values in this column
```

Filling by Exploring Correlations

- An alternative to get less biased estimators for unknowns:
 - to explore the relationships between variables

```
> cor(algae[, 4:18], use="complete.obs") #disregard obs with NAs
```

```
> symnum(cor(algae[, 4:18], use="complete.obs")) #Symbolically encode a given numeric or logical vector or array
```



NH4 and NO3 are positively correlated (0.72)

PO4 and oPO4 are highly correlated (above 0.9)

According to the domain expert, this was expected because the value of the total PO4 includes the value of oPO4

We will find the form of the linear correlation between these variables.

How to Find Linear Relationship



- Find linear relationship between PO_4 and oPO_4

```
> data(algae)
> algae <- algae[-manyNAs(algae), ]
> lm(PO4 ~ oPO4, data=algae)
```

Call:

```
lm(formula = PO4 ~ oPO4, data = algae)
```

Coefficients:

(Intercept)	oPO_4
42.897	1.293

The linear model we have obtained is $\text{PO}_4 = 42.897 + 1.293 \times \text{oPO}_4$

- With this formula, we can fill in the unknown values of these unknowns, provided they are **not** both unknown.
 - Remove the observations with both unknown (sample 62, 199)
 - We have a single observation with an unknown value on PO_4 (sample 28)

Use the Linear Model to Predict



- Use $\text{PO}_4 = 42.897 + 1.293 \times \text{oPO}_4$ to predict the unknown PO_4 at sample 28

```
> algae[28, 'PO4'] <- 42.897 + 1.293 * algae[28, 'oPO4']  
> algae[28,]  
  season  size speed mxPH mnO2 Cl  NO3 NH4 oPO4      PO4 Chla    a1   a2   a3   a4   a5   a6   a7  
28 autumn small  high  6.8 11.1  9 0.63  20      4 48.069  2.7 30.3 1.9  0   0 2.1 1.4 2.1
```

- This can be generalised to fill all missing PO_4 values (if any)

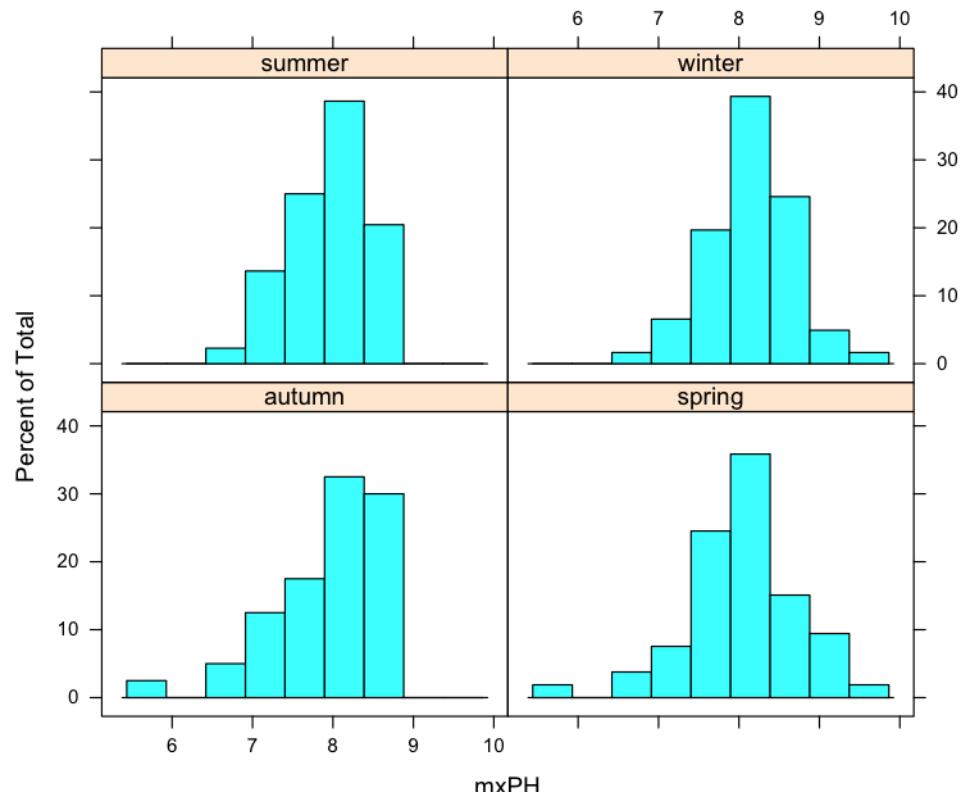
```
> data(algae)  
> algae <- algae[-manyNAs(algae), ]  # delete both unknowns  
> fillPO4 <- function(oP) {  
  if (is.na(oP))  
    return(NA)          #if oPO4's value not available  
  else  
    return(42.897 + 1.293 * oP) #else return the result derived by linear model  
}  
> algae[is.na(algae$PO4), 'PO4'] <-  
  sapply(algae[is.na(algae$PO4), 'oPO4'], fillPO4)  
#This function is applied to all samples with unknown value on the variable PO4
```

Filling by Exploring Correlations

- For other observations with unknown values, we can explore the correlations between the variables and the nominal variables of this problem.

- E.g., mxPH and season

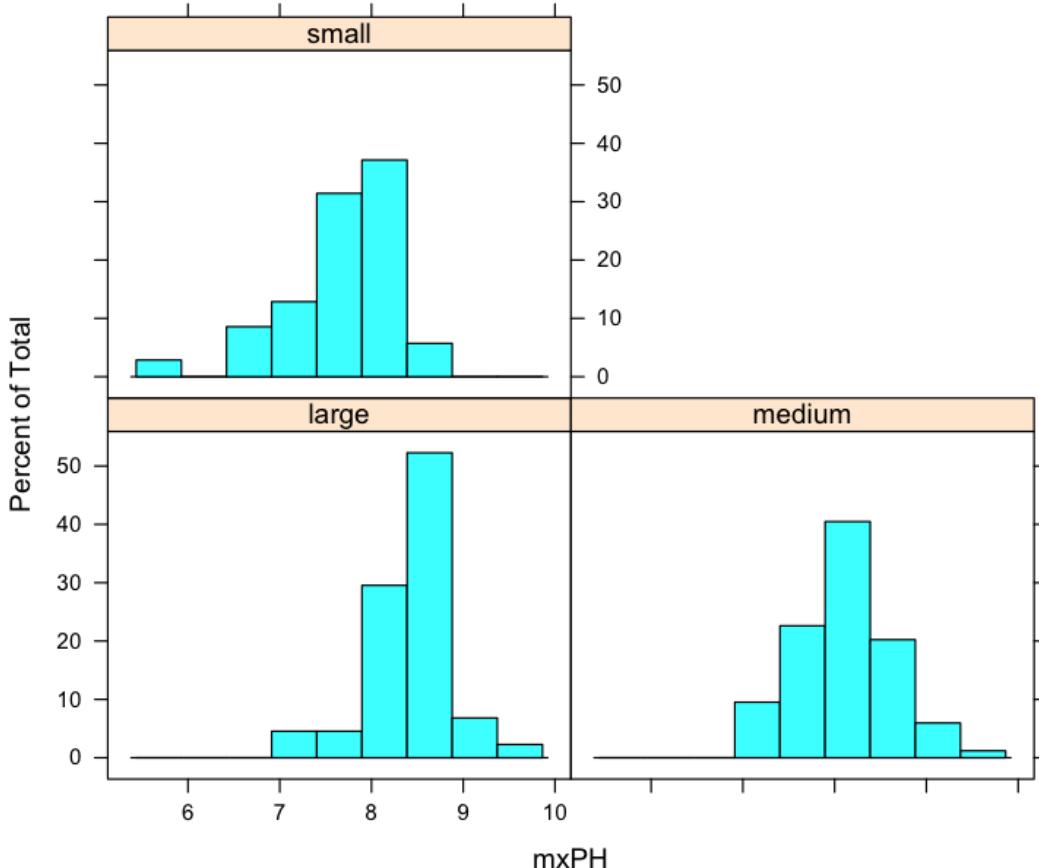
```
> histogram(~ mxPH | season, data=algae)
```



The values of mxPH are not seriously influenced by the season of the year when the samples were collected.

Filling by Exploring Correlations

- If we try the same using the size of the river
 - > `histogram(~ mxPH | size, data=algae)`



What tendency can you observe?

Filling by Exploring Similar Cases



- Another alternative is to use the similarities between the rows to fill in the unknown values
 - If two water samples are similar, and one of them has an unknown value
 - It's very probable that this value is similar to the value of the other sample
 - How to define distance? Which distance can you think of?
 - Euclidean distance!
 - Approach:
 - Find ten most similar cases of any water sample with some unknown value
 - Use their values to fill in the unknown
 - The median of the values of the ten nearest neighbours
 - > `algae <- knnImputation(algae, k=10, meth='median')`
 - The weighted average of the values of the neighbours
 - » The further a neighbour is, the less weight it has (usual weight: $1/d$)
 - > `algae <- knnImputation(algae, k=10) # in DMwR package`

Obtaining Prediction Models

Multiple Linear Regression

Regression Trees

Multiple Linear Regression



- The implementation of linear regression in R is not able to use datasets with unknown values
 - Use the knn-preprocessed technique to fill in the unknowns.

```
> data(algae)
> algae <- algae[!manyNAs(algae), ]
> clean.algae <- knnImputation(algae, k = 10)
```

- Multiple linear regression

```
> lm.a1 <- lm(a1 ~ ., data=clean.algae[,1:12]) # here consider a1 with other 11 predictors
.....
```

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	42.942055	24.010879	1.788	0.07537 .
seasonspring	3.726978	4.137741	0.901	0.36892
seasonsummer	0.747597	4.020711	0.186	0.85270
seasonwinter	3.692955	3.865391	0.955	0.34065
sizemedium	3.263728	3.802051	0.858	0.39179
sizesmall	9.682140	4.179971	2.316	0.02166 *
speedlow	3.922084	4.706315	0.833	0.40573
speedmedium	0.246764	3.241874	0.076	0.93941

Nominal variables are encoded by dummy variables

Erh, where are seasonautumn, sizelarge and speedhigh?

Measures Explained



Coefficients:

	Estimate	Std. Error	t value	Pr(> t) ← p value
(Intercept)	42.942055	24.010879	1.788	0.07537 .
.....				
mxPH	-3.589118	2.703528	-1.328	0.18598
mno2	1.052636	0.705018	1.493	0.13715
Cl	-0.040172	0.033661	-1.193	0.23426
<u>NO3</u>	<u>-1.511235</u>	<u>0.551339</u>	<u>-2.741</u>	<u>0.00674 **</u>
NH4	0.001634	0.001003	1.628	0.10516
<u>PO4</u>	<u>-0.005435</u>	<u>0.039884</u>	<u>-0.136</u>	<u>0.89177</u>
PO4	-0.052241	0.030755	-1.699	0.09109 .
Chla	-0.088022	0.079998	-1.100	0.27265

Signif. codes:	0 '***'	0.001 '**'	0.01 '*'	0.05 '.'
	0.1 ' '	1		

- t test: to see whether each coefficient is statistically significant (hypothesis $H_0: \beta_i=0$)
- $Pr(>|t|)$: a value 0.0001 means that we are 99.99% confident that the coefficient is not null
 - Large value → insignificant factor, small value → significant factor (notice those with *'s by R)

Measures Explained



- R^2 coefficients (multiple and adjusted)
 - Degree of fit of the model
 - pve: proportion variance explained (the smaller, the lack of fit)
 - The adjusted coefficient is more demanding, as it takes into account the number of parameters in the model

Multiple R-squared: 0.3731, Adjusted R-squared: 0.3215

- F-statistics and p-value
 - To test $H_0: \beta_1 = \beta_2 = \dots = \beta_m = 0$
(the target variable does not depend on any of the predictors)
 - p-value: 0.0001 means that we are 99.99% confident that the null hypothesis is not true.
 - If p value is too high (>0.1), it makes no sense to look at the t-test on individual coefficients

F-statistic: 7.223 on 15 and 182 DF, p-value: 2.444e-12

Simply the Linear Model



- Some predictors have a small significance, we could eliminate them from the model

```
> anova(lm.a1) # ANOVA: analysis of variance
```

Analysis of Variance Table

Response: a1

	Df	Sum Sq	Mean Sq	F value	Pr(>F)						
season	3	85	28.2	0.0905	0.9651944						
size	2	11401	5700.7	18.3088	5.69e-08 ***						
speed	2	3934	1967.2	6.3179	0.0022244 **						
mxPH	1	1329	1328.8	4.2677	0.0402613 *						
mnO2	1	2287	2286.8	7.3444	0.0073705 **						
C1	1	4304	4304.3	13.8239	0.0002671 ***						
NO3	1	3418	3418.5	10.9789	0.0011118 **						
NH4	1	404	403.6	1.2963	0.2563847						
oPO4	1	4788	4788.0	15.3774	0.0001246 ***						
PO4	1	1406	1405.6	4.5142	0.0349635 *						
Chla	1	377	377.0	1.2107	0.2726544						
Residuals	182	56668	311.4								

Signif. codes:	0	'***'	0.001	'**'	0.01	'*'	0.05	'.'	0.1	' '	1

It will give us the reduction in the residual sum of squares when adding each variable in turn

season contributes the least to the reduction of the fitting error of the model

Update the Model

- Remove season from the model

```
> lm2.a1 <- update(lm.a1, . ~ . - season)
> summary(lm2.a1)
```

Call:

```
lm(formula = a1 ~ size + speed + mxPH + mnO2 + Cl + NO3 + NH4 +
    OPO4 + PO4 + Chla, data = clean.algae[, 1:12])
```

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	44.9532874	23.2378377	1.934	0.05458 .

.....

speedmedium	-0.2976867	3.1818585	-0.094	0.92556
mxPH	-3.2684281	2.6576592	-1.230	0.22033
mnO2	0.8011759	0.6589644	1.216	0.22561
Cl	-0.0381881	0.0333791	-1.144	0.25407

.....

Multiple R-squared: 0.3682, Adjusted R-squared: 0.3272

F-statistic: 8.984 on 12 and 185 DF, p-value: 1.762e-13

This fit has improved a bit,
but still not too impressive

Previously, adjusted R-squared: 0.3215

Further AVONA Analysis



- Comparison between the two models

```
> anova(lm.a1, lm2.a1)
Analysis of Variance Table
```

Model 1: a1 ~ season + size + speed + mxPH + mnO2 + Cl + NO3 + NH4 + oPO4 + PO4 + Chla

Model 2: a1 ~ size + speed + mxPH + mnO2 + Cl + NO3 + NH4 + oPO4 + PO4 + Chla

Res.Df	RSS	Df	Sum of Sq	F	Pr(>F)
1 182	56668				
2 185	57116	-3	-447.62	0.4792	0.6971

The second model is better, as it has a smaller sum of squares

However, with $\text{Pr}(>F)=0.6971$, it means that only with around 30% confidence we can say the two models are different

- In other words, the difference between the two models are not significant
- The second model is simpler

Automatic Model Simplification



- The `step` function will show you how to simplify the linear model step by step

```
> final.lm=step(lm.a1)
Start: AIC=1152.03          #AIC stands for Akaike Information Criterion
a1 ~ season + size + speed + mxPH + mnO2 + Cl + NO3 + NH4 + oPO4 + PO4 + Chla
```

#omit several steps in the middle, the last step is:

```
Step: AIC=1140.38          # step function use AIC to perform model search
a1 ~ size + mxPH + Cl + NO3 + PO4
```

	Df	Sum of Sq	RSS	AIC
<none>		58517	1140.4	
- mxPH	1	784.1	59301	1141.0
- Cl	1	835.6	59353	1141.2
- NO3	1	1987.9	60505	1145.0
- size	2	2664.3	61181	1145.2
- PO4	1	8575.8	67093	1165.5

AIC offers an estimate of the **relative information lost** when a given model is used to represent the process that generated.

It tells nothing about the absolute quality of a model, only the quality relative to other models. Thus, if all the candidate models fit poorly, AIC will not give any warning of that.

Analyse the Final Model



```
> summary(final.lm)
```

Call:

```
lm(formula = a1 ~ size + mxPH + Cl + NO3 + PO4, data = clean.algae[, 1:12])
```

Residuals:

Min	1Q	Median	3Q	Max
-28.874	-12.732	-3.741	8.424	62.926

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	57.28555	20.96132	2.733	0.00687 **
sizemedium	2.80050	3.40190	0.823	0.41141
sizesmall	10.40636	3.82243	2.722	0.00708 **
mxPH	-3.97076	2.48204	-1.600	0.11130
Cl	-0.05227	0.03165	-1.651	0.10028
NO3	-0.89529	0.35148	-2.547	0.01165 *
PO4	-0.05911	0.01117	-5.291	3.32e-07 ***

Signif. codes:	0 ****	0.001 ***	0.01 **	0.05 *
	'.'	0.1 '	'	1

Residual standard error: 17.5 on 191 degrees of freedom

Multiple R-squared: 0.3527, **Adjusted R-squared: 0.3324**

F-statistic: 17.35 on 6 and 191 DF, p-value: 5.554e-16

In multiple regression, the proportion of variance explained (PVE) is equal to (adjusted) R^2 .

The PVE is still not very interesting (0.3324).

A sign that linearity assumption of this model is inadequate for the domain.

Obtaining Prediction Models

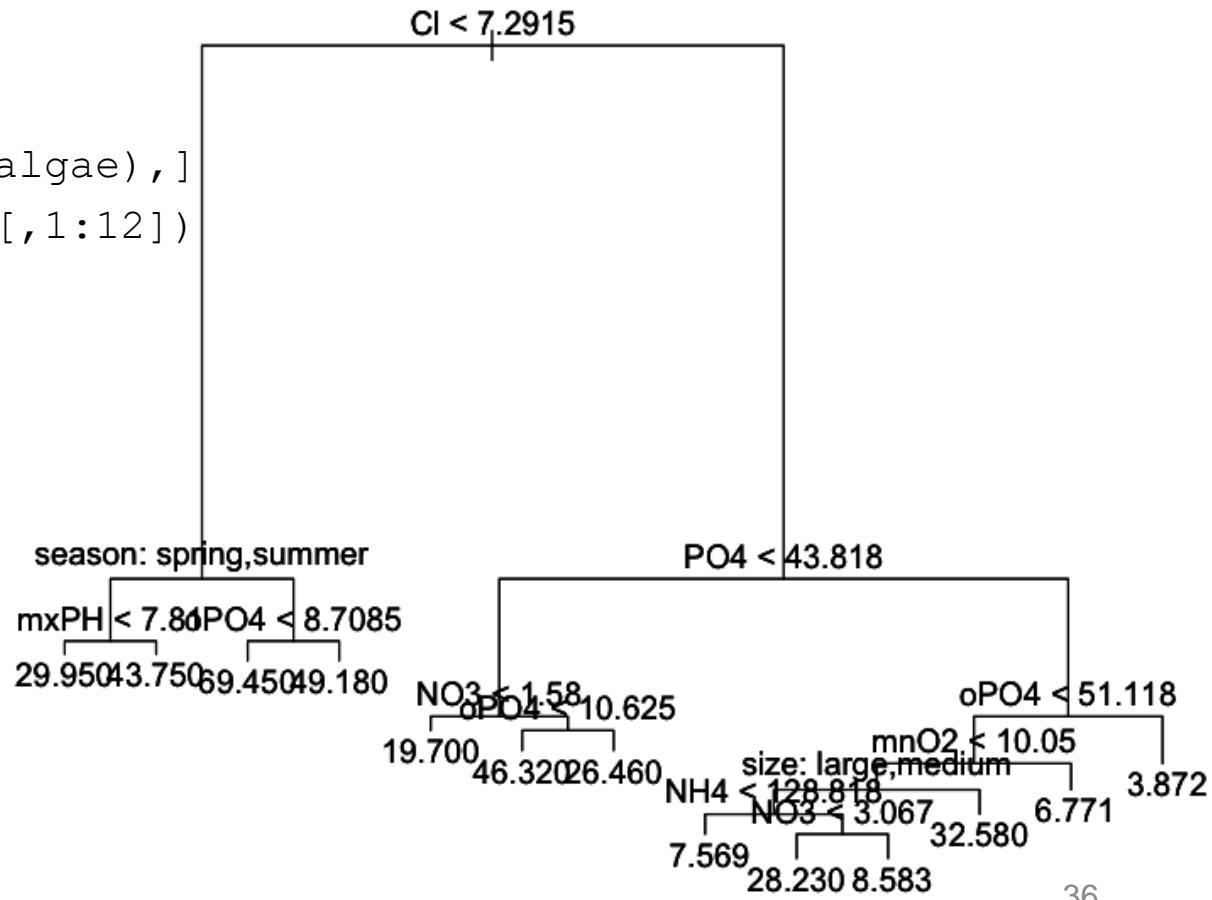
Multiple Linear Regression

Regression Trees

Build a Regression Tree

- It can be done in the same way as building a classification tree

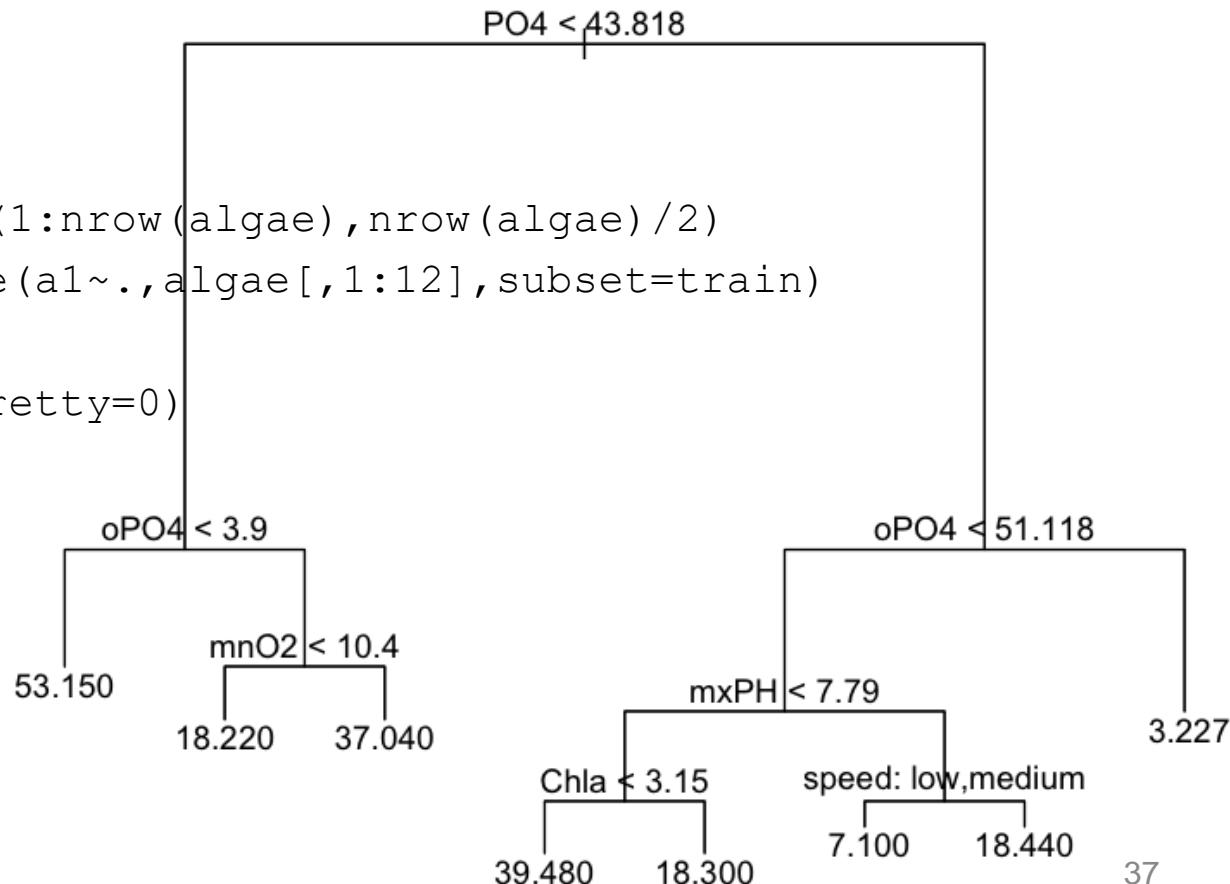
```
> library(tree)
> data(algae)
> algae<-algae[-manyNAs(algae),]
> rt.a1<-tree(a1~.,algae[,1:12])
> text(rt.a1,pretty=0)
```



Build the Tree using Train Part

- Now we randomly sample a train set and build the regression tree based on the set

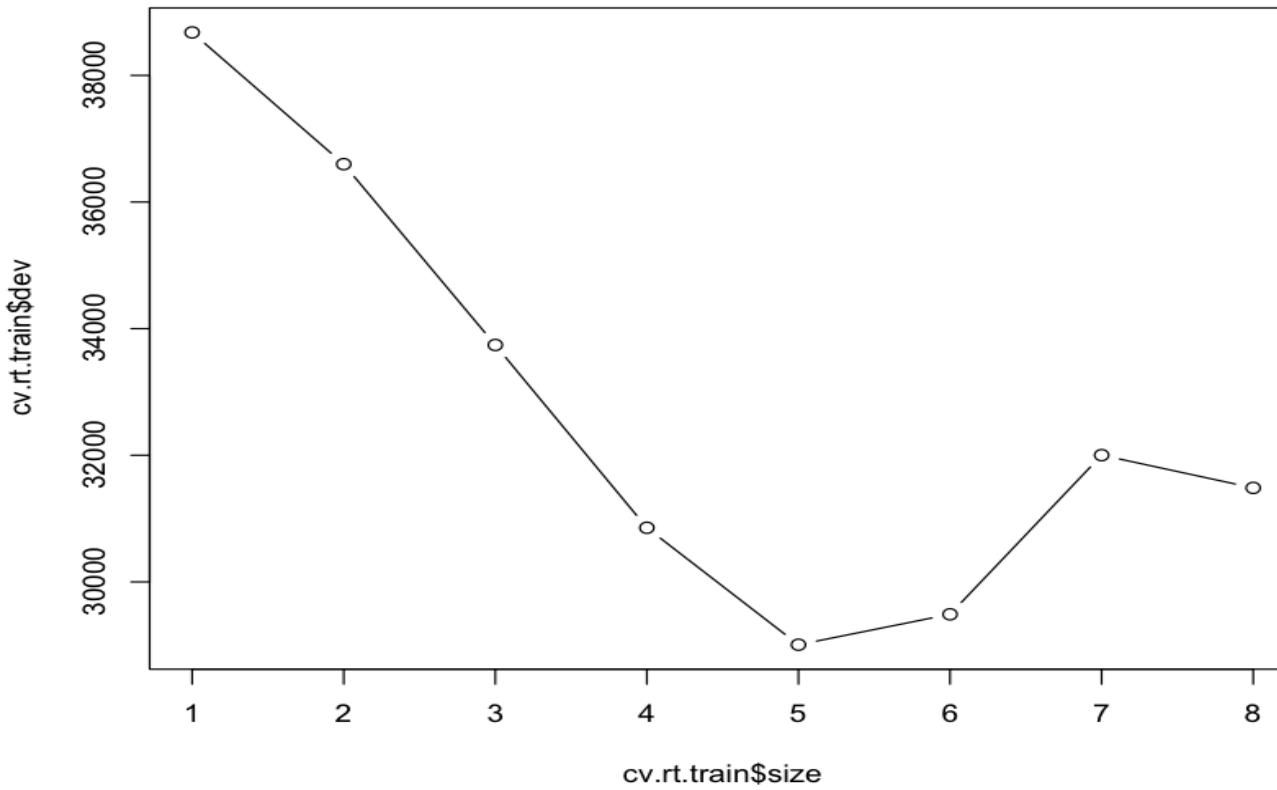
```
> nrow(algae)
[1] 198
> set.seed(2)
> train.al1 <- sample(1:nrow(algae), nrow(algae)/2)
> rt.al1.train <- tree(a1~.,algae[,1:12],subset=train)
> plot(rt.al1.train)
> text(rt.al1.train,pretty=0)
```



Use CV to Check Whether to Prune

- Cross validation is used to see whether the tree `rt.al.train` needs to be pruned

```
> cv.rt.train <- cv.tree(rt.al.train)  
> plot(cv.rt.train$size, cv.rt.train$dev, type='b')
```

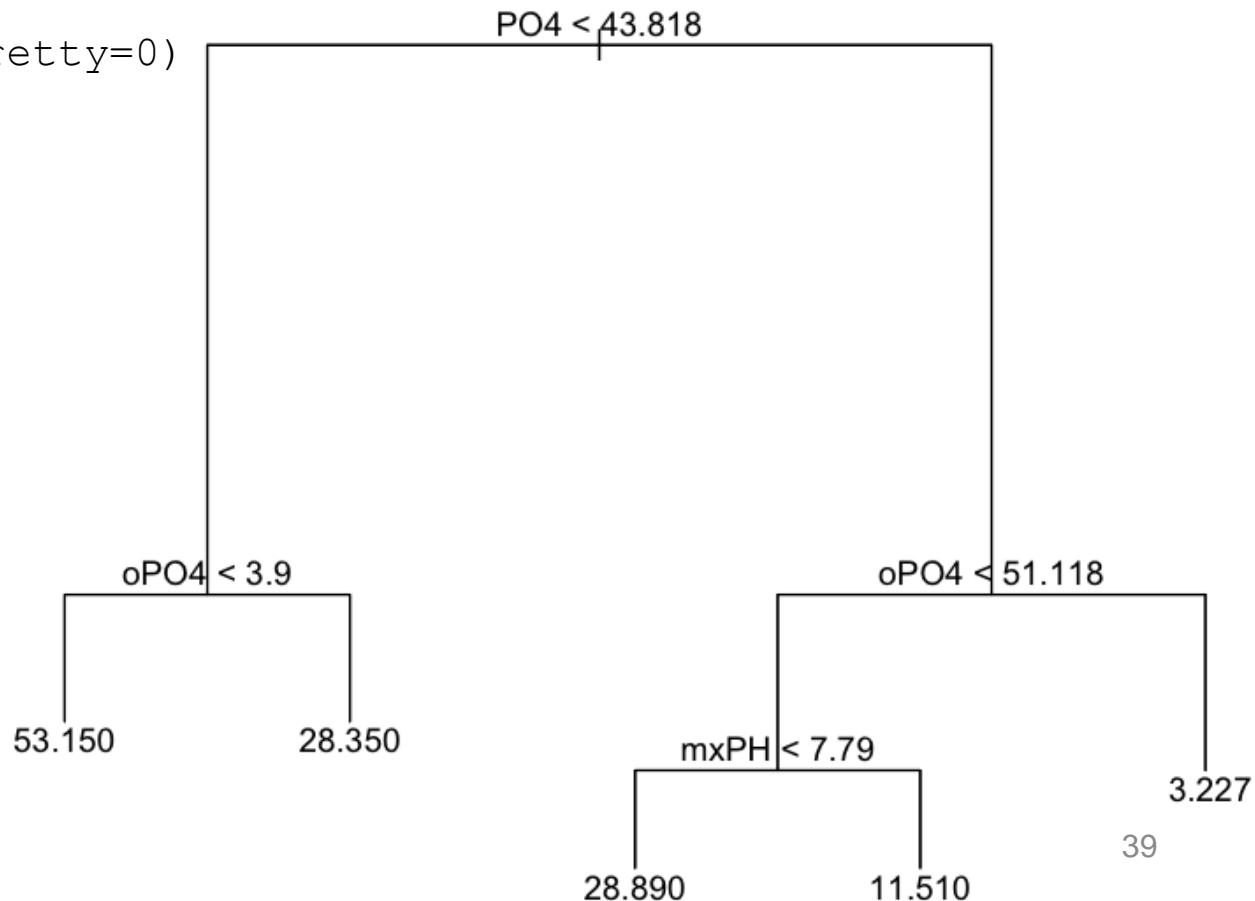


The best tree
(the one with
the minimum
MSE) is of the
size 5

Prune the Tree

- Prune the tree to be of size 5:

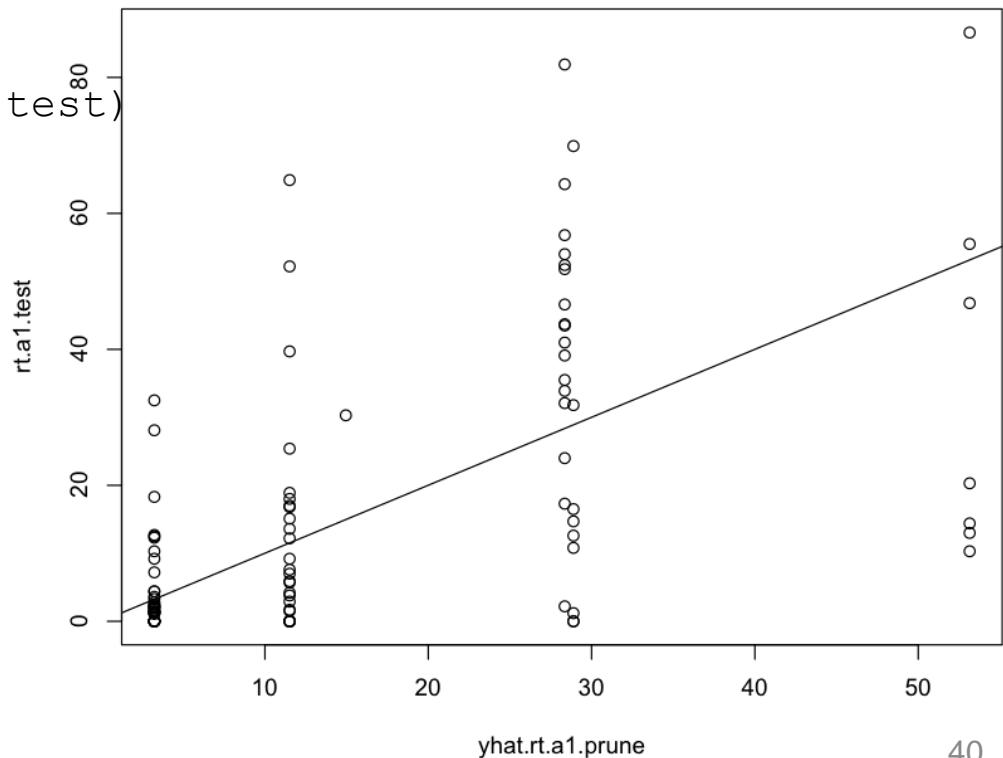
```
> prune.rt.al <- prune.tree(rt.al.train,best=5)  
> plot(prune.rt.al)  
> text(prune.rt.al,pretty=0)
```



Performance Evaluation – Regression Tree

- We use the test part to evaluate the performance

```
> rt.a1.test <- algae[-train,"a1"]  
> yhat.rt.a1.prune <- predict(prune.rt.a1,newdata=algae[-train,1:12])  
> mean((yhat.rt.a1.prune-rt.a1.test)^2)  
[1] 297.0548  
> plot(yhat.rt.a1.prune,rt.a1.test)  
> abline(0,1)
```



Using Bagging

- Since the bagging/randomForest method requires no missing values, we start from the dataset clean.algae

```
> set.seed(20)
> bag.train <- sample(1:nrow(clean.algae), 99)
> bag.al.train <- randomForest(a1~.,clean.algae[1:12],
                                 subset=bag.train,mtry=11,importance=T)
> bag.al.train
```

Call:

```
randomForest(formula = a1 ~ ., data = clean.algae[, 1:12], mtry = 11,
              importance = T, subset = bag.train)
  Type of random forest: regression
  Number of trees: 500
```

No. of variables tried at each split: 11

Mean of squared residuals: 271.161
% Var explained: 42.9

This PVE is
larger than the
linear model⁴¹

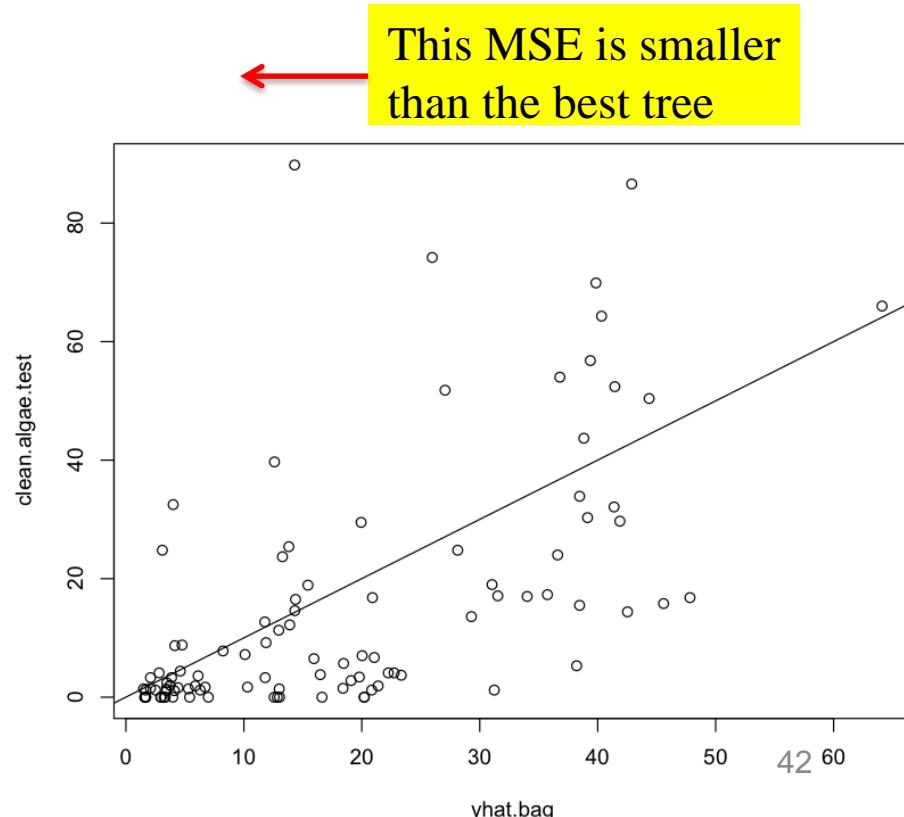
Performance Evaluation - Bagging

- How well does this bagged model perform on the test set?

```
> yhat.bag <- predict(bag.a1.train,newdata=clean.algae[-bag.train,1:12])
> clean.algae.test <- clean.algae[-bag.train,"a1"]
> mean((yhat.bag-clean.algae.test)^2)
[1] 279.9227
> plot(yhat.bag,clean.algae.test)
> abline(0,1)
```

This looks better than in the regression tree →

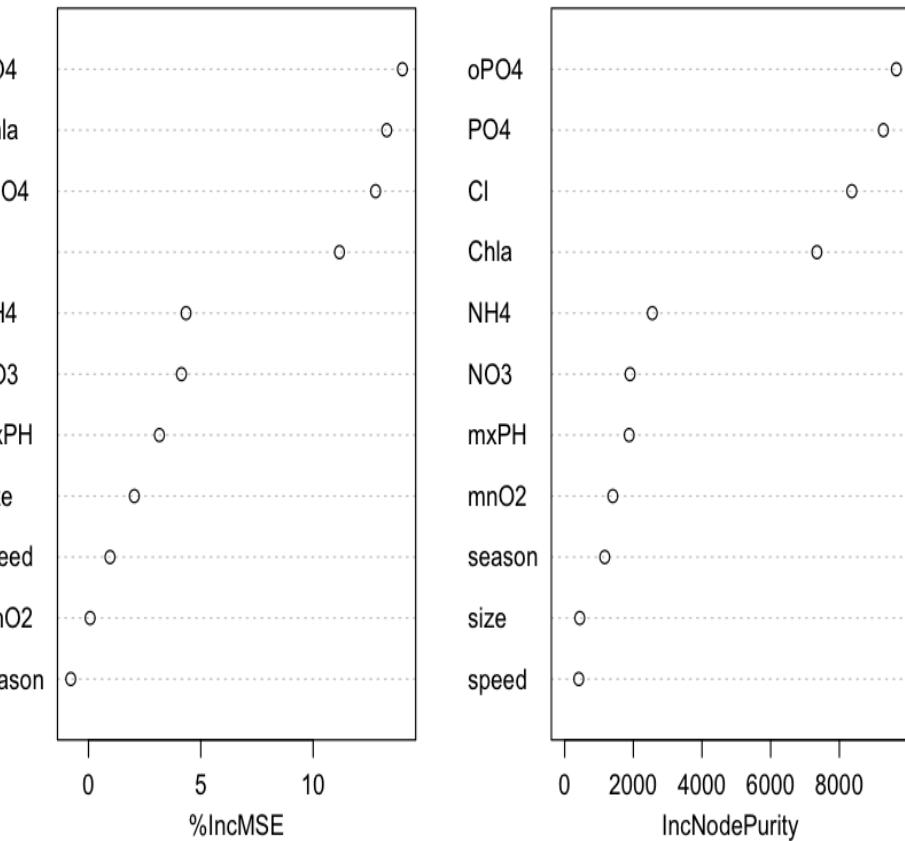
You may play with the number of trees in the bagging at home (ntree=i)



Which Predictors are Important?

```
> importance(bag.a1.train)
      %IncMSE IncNodePurity
season -0.78908700    1168.1408
size    2.04011486    439.9201
speed   0.95764449    410.7687
mxPH    3.15721118    1878.4553
mnO2    0.07241614    1402.3103
Cl      11.17164253   8361.5771
NO3     4.13772152    1904.3340
NH4     4.34200936    2552.3389
oPO4    12.77819198   9659.2113
PO4     13.97331416   9281.1592
Chla   13.27516985   7345.5980
```

```
> varImpPlot(bag.a1.train)
```



Using Random Forest



- Choose a smaller mtry value, usually $p/3$ when building a random forest for regression trees
 - $mtry = 11/3 \approx 3$ or 4

```
> set.seed(20)
> rf.a1.train.4 <- randomForest(a1~.,clean.algae[,1:12], subset=bag.train,
                                    mtry=4, importance=T)
> yhat.rf <- predict(rf.a1.train.4,newdata=clean.algae[-bag.train,1:12])
> mean((yhat.rf-clean.algae.test)^2)
[1] 273.3071
```

mtry = 3 is slightly better
You may find the best mtry at home

```
> set.seed(20)
> rf.a1.train.3 <- randomForest(a1~.,clean.algae[,1:12],subset=bag.train,
                                    mtry=3,importance=T)
> yhat.rf <- predict(rf.a1.train.3,newdata=clean.algae[-bag.train,1:12])
> mean((yhat.rf-clean.algae.test)^2)
[1] 272.3034
```

The PVE of rf.a1.train.3 is 49.6% (use `summary()`), still not very fit

Probably try nonlinear models (polynomials, etc), something for you to try at home, too

Prediction for New Test Set

Prediction for the Algae



- We are given 140 test samples, whose algae levels are unknown.
- We will choose the best models to obtain these predictions.
 - To obtain unbiased estimates of MSE for a set of models
 - By means of a cross-validation experimental process
 - For simplicity, we only predict a_1
- For a_1 , we have already shown that the randomForest model `rf.a1.train.3` is the best model
 - Use `rf.a1.train.3` to make the prediction

Unknowns in the Test Data



- There are unknowns in the test data
- We could use `knnImputation()` as in the training dataset
 - Use other test cases to fill in the unknowns → not ideal
 - Use training data to find the neighbours instead
 - use `knnImputation()`, but with an extra argument

```
> clean.test.algae <- knnImputation(test.algae, k=10, distData=algae[,1:11])
```

The `distData` argument allows you to supply an extra set of data (i.e., the training dataset) where the ten nearest neighbours are to be found for each case with unknowns in the `test.algae` dataset.

Make the Prediction



- Finally,...

```
> preds <- rep(0,140)
> preds <- predict(rf.al.train.3, newdata=clean.test.algae, mtry=3, importance=T)
> preds
   1          2          3          4          5          6          7          8          9
7.266943 10.458083 13.387457 13.542400 27.145823 33.591357 35.073133 37.611920 38.065740
      10         11         12         13         14         15         16         17         18
36.190503 10.706703 15.288940 40.884010 38.163287 37.630820 26.044157 10.487700 20.337720
      19         20         21         22         23         24         25         26         27
40.361043 54.538080 6.965607 4.724927 4.981443 11.896803 6.452217 5.023043 24.228200
      28         29         30         31         32         33         34         35         36
43.114077 27.373763 23.633090 26.444843 20.911110 32.294507 38.157100 55.714590 35.624243
      37         38         39         40         41         42         43         44         45
35.052197 51.597240 33.467427 39.437900 37.612970 16.618960 10.317370 9.975300 10.411817
      46         47         48         49         50         51         52         53         54
3.337300 10.015007 5.438577 17.838527 31.355300 11.017717 3.678907 5.509753 3.913507
      55         56         57         58         59         60         61         62         63
4.779007 12.729870 13.189073 11.902373 17.185123 14.290100 6.853557 21.050563 16.727573
      64         65         66         67         68         69         70         71         72
8.964617 33.982597 27.070277 18.403937 40.085983 43.577550 4.610323 6.584670 4.338993
.....
```



WORK
HARD
AND
WAIT FOR
CHRISTMAS